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ABSTRACT

Bone Morphogenetic Protein 15 (BMP 15) is a member of transforming growth factor- β (TGF- β) superfamily which plays an important role in ovarian follicular development and ovulation rate. In this study a partial sequence of this gene was screened for polymorphisms. PCR amplification of DNA isolated from blood of 100 Egyptian buffaloes hiefers (50 infertile and 50 normal). Revealed two loci, BMP15.1 (222 bp) and BMP15.2 (222 bp) which includes exon 2 of BMP 15 gene. Polymorphisms in these two loci were detected by using direct sequencing and single strand conformational polymorphism (SSCP) technique. A single pattern of SSCP was detected. This was confirmed by nucleotide sequencing which revealed absence of single nucleotides polymorphisms (SNPs) or any other type of polymorphisms in the two loci. The sequencing and PCR-SSCP patterns revealed no polymorphism in all studied animals. Further studies must be done on another loci on BMP 15 gene or other genes related to infertility in Egyptian buffaloes.

Keywords: BMP15 gene, polymorphism, Egyptian buffalo, infertility, PCR-SSCP.

INTRODUCTION

Buffaloes belongs to Bovidae family and Ruminantia suborder. Egyptian buffalo had classified geographically and some phenotypic differences into Beheri, Baladi, Menoufi and Saidi *(FAODAD-IS, 2013)*. The domestic water buffalo has a great economic importance for small farmers in developing countries like Egypt *(Michelizzi et al., 2010)*.

Infertility defined as the loss of ability to conceive after one year of regular mating or insemination, although, in many cases, the diagnosis is unexplained, several causes such as decreased ovulation rate, mechanical stoppage, deficiencies of sperm and parental age. Repeat breeder is the main source of economic loss in dairy farms. Infertility genetically may result from gene disorders, abnormalities in chromosomes, and phenotypes with multifactorial inheritance (Sah et al., 2003).

Transforming growth factor was firstly identified as a secreted factor which control cell proliferation *(Roberts and Sporn, 1990).* Based on cell signaling pathways, the proteins

can be classified into two major groups, the Morphogenetic Proteins/Growth Bone Differentiation Factors. and activins/Transforming Growth Factors (Chang et al., 2002). Several BMP family members, including BMP -2, 3, 4, 5, 6, 7, and 15 are expressed in uterus (Tanwar and McFarlane, 2011), ovary (Sun et al., 2010) and in the granulosa cells and oocyte (Otsuka et al., 2001). They perform great biological activities as regulators in development of ovarian follicles. female reproductive tract differentiation, blastocyst implantation in the uterus, and organogenesis and morphogenesis during embryo development (Kishigami and Mishina, 2005).

The aim of the present study was to search for polymorphisms in *BMP15 gene* in infertile and normal buffalo heifers using PCR –SSCP and DNA sequencing.

MATERIAL AND METHODS

All experiments were conducted in the Biotechnology Lab, Department of Animal Wealth Development, Faculty of Veterinary Medicine, Zagazig University, Egypt.

2.1. Animals and sample collection

The experiments were performed in 2014–2015 from a buffalo nucleus herd kept in Nataff-Kadeem Station and Mahalet-Mousa Farm, Animal Production Research Institute (APRI). A total of 100 heifers (50 repeat breeders and 50 normal) were genotyped for the *BMP15* gene. Blood samples were obtained from jugular vein into sterilized tubes

containing EDTA as an anticoagulant and stored at -20° C until use.

2.2. Genomic DNA extraction

Total genomic DNA was extracted using DNAeasy Blood & Tissue Kit (Fermentas, Thermo scientific, #K0721) according to the manufacturer's protocol.

2.3. PCR amplification

Two loci of BMP15 gene were amplified by PCR, BMP15.1 and BMP15.2 which includes in exon 2. The PCR reactions were performed with a total volume 20 µL containing 2 µL DNA template, 1 µL from each primer, 10 µL master mix (Thermo Scientific, #K1071) and 6 µL nuclease free water. This mix was put in thermal cycler and condition was optimized as shown in table 1. PCR products were electrophoresed on 1.5% agarose gels. Fragment size was determined using Gene Ruler[™] 100bp Ladder (Fermentas).

2.4. Genotyping using PCR-SSCP technique

PCR-SSCP was performed as previously described by *El-Magd et al*, (2014) with minor modification.

2.5. DNA sequencing

The purified PCR products were sequenced by automated sequencer (ABI 310, Applied Biosystem, USA). The Sequences were analyzed using geneious 4.8.4 software. The obtained sequences were edited manually using CLC Main Workbench 7 to identify nucleotide substitutions.

RESULTS

The amplified product of the *BMP 15.1* locus was 222 bp (Fig 1) and that of *BMP 15.2* locus was 222 bp (Fig 2). Analysis of SNPs in Exon 2 of BMP15 gene using PCR-SSCP revealed no SNPs were detected in either *BMP15.1* locus (Fig 3) or *BMP15.2* locus (Fig 4). Nucleotide sequencing confirmed the

results and showed no SNPs in the two loci of BMP15 gene (Figs. 5, 6).

To detect polymorphic sites/SNPs, all sequenced amplicons of *BMP15* were aligned and compared using Clustal Omega and CLC Main Workbench7 software. The results revealed that there are no differences between all studied (infertile and normal animals) in all sequences of BMP15.1 locus and BMP15.2 locus (**Fig 7, 8**).

Table (1): Primer sequences and PCR condition of *BMP15* gene.

Locus	primer	PCR condition			
BMP15.1	F 5 GAGTGTTCAGAAGACCAAACCTC3	94 °C /3 min, 35 cycles (94			
	R 5 TGGGGAGCAATGATCCAGTGATCC3	°C for 30 s ,60°C for 40 s, 72 °C for 1 min) and final			
BMP15.2	F 5 CTACTGTAAGGGAGTATGTCCTCG3	extension at 72 °C for 5			
	R Ś CTGCATGTGCAGGACTGGGCAAŚ	min.			



Figure (1): Ehidium bromide stained agarose gel of PCR product representing amplification of *BMP15.1* (222 bp) from normal and infertile buffalo heifers. M: 100 bp ladder.

1000 bp M									
222 bp 1000 bp	-	-	-	-	-	-	-	-	-

Figure (2): Ehidium bromide stained agarose gel of PCR product representing amplification of *BMP 15.2* (222 bp) from normal and infertile buffalo heifers. M: 100 bp ladder.

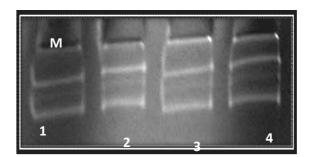


Figure (3): PCR-SSCP patterns of BMP 15.1 locus in Egyptian buffalo show one trimorphic SSCP pattern was detected.

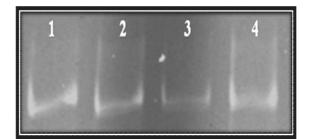


Figure (4): PCR-SSCP patterns of BMP 15. 2 locus in Egyptian buffalo show one dimorphic SSCP pattern.

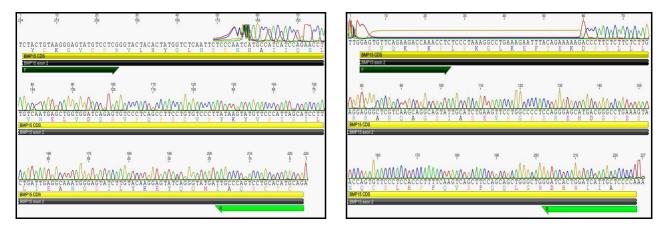


Fig (5). Nucleotide sequences of BMP15.1 locus Fig (6). Nucleotide sequences of BMP15.2 locus taken as representative example.

taken as representative example.

238

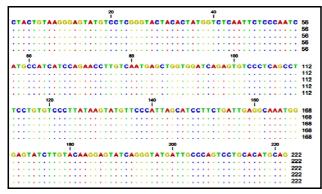


Fig (7). DNA sequence alignment of BMP 15.1 Fig (8). DNA sequence alignment of BMP 15.2 locus locus.

61 122 122 122 122 122 183 183 183 183 183

DISCUSSION

The discovery of molecular biology and DNA based markers created new methods to study livestock genetics and animal breeding. Selection which depends on genotype can enhance productivity of farm animals, also enhance the adaptation to environment and keep on genetic diversity (Haves, 2007). Molecular markers considered one of the most vital tools for the genome analysis and enable association of heritable traits with the underlying genomic variation. Also it can studies of candidate genes and their effect on the phenotypic traits are the basis for Marker assisted selection (Kulibaba and PodStreshnyi, 2012).

Previous studies have revealed that spontaneous mutations in the ovine and murine BMP15 (FecX) gene may alter fertility and ovulatory rate in these species. Infertile animals with arrested folliculogenesis at primary stage results from deletion of the BMP15 (Hanrahan et al., 2004).

agree with the finding This of Gholibeikifard et al., (2014) whose used PCR-SSCP technique and showed no evidence of mutation was observed in BMP15 gene in

Iranian Bluchi Sheep Breed, all of which were monomorphic for exon 2 BMP-15 gene.

Zhang et al., (2009) calculated the allele and genotype frequencies of the BMP15 gene in 8 breeds of cattles are presented two genotypes (AA and AB) were detected, while only one genotype (AA) was detected in Menggolian, Holstein and Simmental, this result disagrees with our result.

Some mutations in exon 2 of BMP15 gene have been associated with prolificacy in various breeds of sheep. Polymorphism of BMP15 gene exon 2 was studied in 100 Mehraban and 100 Lori ewes, using PCR-SSCP and DNA sequencing methods. A new point mutation $(G \rightarrow A)$ was found at position 57 of the amplified fragment (Zamani et al., 2015).

CONCLUSION

PCR-SSCP failed to produce patterns capable of discriminating between the exon 2 of BMP15 gene in infertile and normal heifers so further studies should be done on other loci on BMP15 or on another gene related to infertility in Egyptian buffalo.

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REFERENCES

- Chang, H., Brown, C.B. and Matzuk, M.M. (2002): Genetic analysis of the Mammalian transforming growth factor_superfamily. Endocrinol. Rev. 23, 787–823.
- El-Magd, M.A., Abo-Al-Ela, H.G., El-Nahas, A., Saleh, A.A., Mansour, A.A. (2014) : Effects of a novel SNP of IGF2R gene on growth traits and expression rate of IGF2R and IGF2 genes in gluteus medius muscle of Egyptian buffalo. Gene 540, 133-139.
- FAODAD-IS, (2013): FAO Domestic Animal Diversity Information System (DAD-IS). http://dad.fao.org/. Access 7 Feb 2013.
- Gholibeikifard, A.; Afshar, M. A.;
 Mashhadi, M. H. and Mohammadi,
 H. (2014): Molecular Study on the Exon 2 Region of the Ovis Bone Morphology Protein 15 (BMP-15)
 Gene in Iranian Bluchi Sheep Breed by PCR-SSCP Technique. Iranian Journal of Applied Animal Science. 4 (4): 769-773.
- Hayes, B. (2007): QTL mapping, MAS, and genomic selection. Iowa State University. Department of Animal Sciences.

- Hanrahan, J.P.; Gregan, S.M.; Mulsant, P.;
 Mullen, M. Davis, G.H.; Powell, R. and Galloway, S.M. (2004):
 Mutations in the genes for oocytederived growth factors GDF9 and BMP15 are associated with both increased ovulation rate and sterility in Cambridge and Belclare sheep (Ovis aries). Biol. Reprod., 70: 900-909.
- Kishigami, S. and Mishina, Y. (2005): BMP signaling and early embryonic patterning. Cytokine Growth Factor Rev. 16:265–78.
- Kulibaba, R.A. and Podstreshnyi, A.P. (2012): Prolactin and growth hormone gene polymorphisms in chicken lines of Ukrainian selection. Cytology and Genetics, 46: 390-395.
- Michelizzi, V.N., Dodson, M.V., Pan, Z.X., Amaral, M.E.J., Michal, J.J., McLean, D.J., Womack, J.E. and Jiang, Z.H. (2010): Water Buffalo Genome Science Comes of Age. International Journal of Biological Sciences. 6, 333-349.
- Otsuka, F., Yamamoto, S., Erickson, G.F. and Shimasaki, S. (2001): Bonemorphogenetic protein-15 inhibits follicle-stimulating hormone (FSH) action by suppressing FSH receptor expression. J. Biol. Chem. 276: 11387-11392.
- Roberts, A.B. Sporn, M.B. (1990): The transforming growth factors. In:Sporn, M.B., Roberts, A.B. (Eds.), Peptide Growth Factors and Their Receptors, Part I. Springer-Verlag, Berlin, pp. 419–472.
- Sah, S.K., Sivapalam, G., Gibbons, N., Tempest, H. and Griffith, D.K. (2003): The genetic Basis of Infertility. Reproduction, 126 (1): 13-25.

- Sun, R.Z., Lei, L., Cheng, L., Jin, Z.F., Zu, S.J., and Shan, Z.Y. (2010): Expression of GDF-9, BMP-15 and their receptors in mammalian ovary follicles. J. Mol. Histol, 41: 325–32.
- Tanwar, P.S. McFarlane, J.R. (2011): Dynamic expression of bone morphogenetic protein 4 in reproductive organs of female mice. Reproduction, 142:573–9.
- Zamani, P.; Nadri, S.; Saffaripour, R.; Ahmadi, A.; Dashti, F. and Abdoli, R. (2015): A new mutation in exon 2

of the bone morphogenetic protein 15 gene is associated with increase in prolificacy of Mehraban and Lori sheep. Trop Anim Health Prod. 47(5):855-60.

Zhang, L.P., Gan Q.F., Zhang X.H., Li H.D., Hou, G.Y., Li J.Y., Gao, X.. Ren, H.Y., Chen, J.B. and Xu, S.Z. (2009): Detecting a deletion in the coding region of the bovine bone morphogenetic protein 15 gene (BMP15). J. Appl Genet. 50 (2): 145– 148.

الملخص العربي

وعلاقته BMP15 تحديد الطفرات الموضعية في الاكسون الثاني للجين

بالعقم في الجاموس المصري

خيرى محمد البيومى"، ايمان السيد العربى"، ايمن عبد اللطيف صالح"، هدى ذكى عثمان"، محمود صلاح الطربانى"، اشرف فتحى عوض"، امير حسن عبد الفتاح"، محمد ابو المجد الغنام"".

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أجريت هذه الدراسة على الجين BMP15 والذى يعتبر عاملا مهما من عوامل النمو التى تلعب دورا كبيرا فى نمو البويضات وتنشيط عملية التبويض. تمت هذه الدراسة على بعض المواقع الجينية التى تمثل الاكسون الثانى حيث تم اخذ عينات دم من ١٠٠ حيوان من الجاموس المصرى (٥٠ حيوان طبيعى و٥٠ حيوان يعانى من عدم الخصوبة) تم اخذ عينات دم من ١٠٠ حيوان من الجاموس المصرى (٥٠ حيوان طبيعى و٥٠ حيوان يعانى من عدم الخصوبة) وتم استخلاص الحامض النووى DNA وتضخيمه باستخدام تفاعل البلمرة المتسلسل PCR ثم تحديد الطفرات الموضعية باستخدام الشريط الفردى SSCP وتاكيده بعمل تتابع نيوكليوتيدى sequencing. اظهرت النتائج عدم وجود طفرات موضعية فى المواقع تحت الدراسة.